

# Package ‘CACIMAR’

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**Title** Cross-Species Analysis of Cell Identities, Markers and Regulations

**Version** 1.0.0

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**Description** A toolkit to perform cross-species analysis based on scRNA-seq data. This package contains 5 main features. (1) identify Markers in each cluster. (2) Cell type annotation (3) identify conserved markers. (4) identify conserved cell types. (5) identify conserved modules of regulatory networks.

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**Encoding** UTF-8

**LazyData** true

**RoxygenNote** 7.1.1

**Imports** pheatmap, pbapply, psych, ROCR, reshape2, dplyr, grDevices, stats, methods, viridisLite

**Depends** R (>= 4.0), Seurat

**Suggests** knitr, testthat (>= 3.0.0)

**Config/testthat/edition** 3

**URL** <https://github.com/jiang-junyao/CACIMAR>

**BugReports** <https://github.com/jiang-junyao/CACIMAR/issues>

**NeedsCompilation** no

**Repository** CRAN

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CACIMAR_cols	<i>CACIMAR colors palette</i>
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## Description

CACIMAR colors palette

## Usage

CACIMAR\_cols(color\_number)

## Arguments

color\_number    numeric, indicating used colors number

## Value

vector of colors

## Examples

```
CACIMAR_cols(10)
CACIMAR_cols(20)
```

---

Format\_Markers\_Frac     *Format marker genes for plotting*

---

**Description**

Order the gene expression in each cluster to make the heatmap look better

**Usage**

```
Format_Markers_Frac(Marker_genes)
```

**Arguments**

Marker\_genes     data.frame, generated by [Identify\\_Markers](#)

**Value**

Markers corresponding to certain cluster

**Examples**

```
data("pbmc_small")
all.markers <- Identify_Markers(pbmc_small)
all.markers2 <- Format_Markers_Frac(all.markers)
```

---

Heatmap\_Cor     *plot the heatmap of marker genes across different species*

---

**Description**

plot the heatmap of marker genes across different species

**Usage**

```
Heatmap_Cor(
  RNA1,
  RowType1 = "",
  ColType1 = "",
  cluster_cols = TRUE,
  cluster_rows = FALSE,
  Color1 = NULL,
  ...
)
```

**Arguments**

RNA1	correlation of expression in each cell type
RowType1	character, indicating the cell types that you want to show on the row in heatmap. RowType1="" means show all cell types
ColType1	character, indicating the cell types that you want to show on the column in heatmap. RowType1="" means show all cell types
cluster_cols	boolean values determining if columns should be clustered or hclust object
cluster_rows	boolean values determining if rows should be clustered or hclust object
Color1	vector of colors used in heatmap
...	parameter in pheatmap

**Value**

pheatmap object

**Examples**

```
load(system.file("extdata", "network_example.rda", package = "CACIMAR"))
n1 <- Identify_ConservedNetworks(OrthG_Mm_Zf,mmNetwork,zfNetwork,'mm','zf')
Heatmap_Cor(n1[[2]],cluster_cols=TRUE, cluster_rows=FALSE)
```

---

Identify\_CellType      *Identify cell type of each cluster*

---

**Description**

This function has three steps to identify cell type of each cluster. (1) Calculate the power of each known marker based on AUC (area under the receiver operating characteristic curve of gene expression) which indicates the capability of marker  $i$  from cell type  $m$  to distinguish cluster  $j$  and the other clusters. (2) Calculate the united power (UP) for cell type  $m$  across each cluster  $j$ . (3) For each cluster  $j$  we determine the cell type according to UP. Generally, the cluster belongs to the cell type which have the highest united power or higher than the threshold of the united power (for example  $> 0.9$  power).

**Usage**

```
Identify_CellType(seurat_object, Marker_gene_table)
```

**Arguments**

seurat_object	seurat object
Marker_gene_table	data.frame, indicating marker gene and its corresponding cell type. Marker_gene_table should contain two columns: 'CellType' represent corresponding cell types of each marker and 'Marker' represent Markers

**Value**

Cell type with the highest power in each cluster

**Examples**

```
KnownMarker=data.frame(c('AIF1', 'BID', 'CCL5', 'CD79A', 'CD79B', 'MS4A6A'),c('a', 'a', 'a', 'b', 'b', 'b'))
data("pbmc_small")
colnames(KnownMarker)=c('Marker', 'CellType')
CT <- Identify_CellType(pbmc_small, KnownMarker)
```

---

**Identify\_ConservedCellTypes**

*Identify conserved cell types based on power of genes and orthologs database*

---

**Description**

Identify conserved cell types based on power of genes and orthologs database

**Usage**

```
Identify_ConservedCellTypes(
  OrthG,
  Species1_Marker_table,
  Species2_Marker_table,
  Species_name1,
  Species_name2
)
```

**Arguments**

OrthG            ortholog genes database

Species1\_Marker\_table  
                 data.frame of species 1, should contain three column: 'gene', 'cluster' and 'power'

Species2\_Marker\_table  
                 data.frame of species 2, should contain three column: 'gene', 'cluster' and 'power'

Species\_name1    character, indicating the species names of Species1\_Marker\_table

Species\_name2    character, indicating the species names of Species2\_Marker\_table

**Value**

list contains two elements: first one is details of conserved cell types, second one is matrix of cell types conserved score

**Examples**

```
load(system.file("extdata", "CellTypeAllMarkers.rda", package = "CACIMAR"))
expression <- Identify_ConservedCellTypes(OrthG_Mm_Zf,mm_Marker[1:30,],zf_Marker[1:30,],'mm','zf')
```

---

Identify\_ConservedMarkers

*Identify orthologs marker genes for two species*

---

**Description**

Identify orthologs marker genes for two species based on orthologs database

**Usage**

```
Identify_ConservedMarkers(
  OrthG,
  Species1_Marker_table,
  Species2_Marker_table,
  Species_name1,
  Species_name2,
  match_cell_name = NULL
)
```

**Arguments**

OrthG                    ortholog genes database

Species1\_Marker\_table                    data.frame of species 1, first column should be gene name, second column should be Clusters corresponding to marker gene

Species2\_Marker\_table                    data.frame of species 2, first column should be gene name, second column should be Clusters corresponding to marker gene of marker genes.

Species\_name1            character, indicating the species names of Species1\_Marker\_table.

Species\_name2            character, indicating the species names of Species2\_Marker\_table

match\_cell\_name                          characters contained in both cell names to match similar cell types

**Value**

Data frame of conserved markers

**Examples**

```
load(system.file("extdata", "CellMarkers.rda", package = "CACIMAR"))
o1 <- Identify_ConservedMarkers(OrthG_Mm_Zf,Mm_marker_cell_type,
Zf_marker_cell_type,Species_name1 = 'mm',Species_name2 = 'zf')
o2 <- Identify_ConservedMarkers(OrthG_Zf_Ch,Ch_marker_cell_type,
Zf_marker_cell_type,Species_name1 = 'ch',Species_name2 = 'zf')
```

---

`Identify_ConservedNetworks`*Identify conserved regulatory networks*

---

**Description**

Use Score of Conserved network to identify conserved regulatory network modules based on homologous genes databased and topology of networks

**Usage**

```
Identify_ConservedNetworks(  
  OrthG,  
  Species1_GRN,  
  Species2_GRN,  
  Species_name1,  
  Species_name2  
)
```

**Arguments**

<code>OrthG</code>	ortholog genes database
<code>Species1_GRN</code>	gene regulatory network of species 1
<code>Species2_GRN</code>	gene regulatory network of species 2
<code>Species_name1</code>	character, indicating the species names of <code>Species1_GRN</code>
<code>Species_name2</code>	character, indicating the species names of <code>Species2_GRN</code>

**Value**

list contains two df. First df contains details of conserved regulatory network, second df contains NCS between module pairs

**Examples**

```
load(system.file("extdata", "gene_network.rda", package = "CACIMAR"))  
n1 <- Identify_ConservedNetworks(OrthG_Mm_Zf,mm_gene_network,zf_gene_network,'mm','zf')
```

---

Identify\_Markers      *Identify markers of each cluster*

---

### Description

This function first identify marker genes in each cluster with Roc threshold  $> \text{RocThr}$ . Then, based on marker genes identified above, this function calculates the difference and power of marker genes in each cluster, and marker genes with Difference threshold  $> \text{DiffThr}$  will be retained. Next, gene with the largest power in which cluster will be the marker gene in this cluster. Eventually, make fisher test for power of each cluster, cluster with  $p.\text{value} < 0.05$  will be retained as the final cluster for marker gene

### Usage

```
Identify_Markers(  
  Seurat_object,  
  PowerCutoff = 0.4,  
  DifferenceCutoff = 0,  
  PvalueCutoff = 0.05  
)
```

### Arguments

Seurat\_object    Seurat object, should contain cluster information

PowerCutoff      numeric, indicating the cutoff of gene power to refine marker genes

DifferenceCutoff      numeric, indicating the cutoff of difference in marker genes between clusters to refine marker genes

PvalueCutoff      numeric, indicating the p.value cutoff of chi-square test to refine marker genes

### Value

Data frame of conserved markers

### Examples

```
data("pbmc_small")  
all.markers <- Identify_Markers(pbmc_small)
```



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OrthG_Hs_Ch	<i>Orthologs genes database for homo sapiens and zebrafish</i>
-------------	--

---

**Description**

Orthologs genes database for homo sapiens and zebrafish

**Usage**

OrthG\_Hs\_Ch

**Format**

An object of class data.frame with 12219 rows and 5 columns.

---

OrthG_Hs_Mm	<i>Orthologs genes database for homo sapiens and mus musculus</i>
-------------	---

---

**Description**

Orthologs genes database for homo sapiens and mus musculus

**Usage**

OrthG\_Hs\_Mm

**Format**

An object of class data.frame with 16754 rows and 5 columns.

---

OrthG_Hs_Zf	<i>Orthologs genes database for homo sapiens and zebrafish</i>
-------------	--

---

**Description**

Orthologs genes database for homo sapiens and zebrafish

**Usage**

OrthG\_Hs\_Zf

**Format**

An object of class data.frame with 12017 rows and 5 columns.

---

OrthG_Mm_Ch	<i>Orthologs genes database for mus musculus and chicken</i>
-------------	--

---

**Description**

Orthologs genes database for mus musculus and chicken

**Usage**

OrthG\_Mm\_Ch

**Format**

An object of class data.frame with 62661 rows and 5 columns.

---

OrthG_Mm_Zf	<i>Orthologs genes database for mus musculus and zebrafish</i>
-------------	--

---

**Description**

Orthologs genes database for mus musculus and zebrafish

**Usage**

OrthG\_Mm\_Zf

**Format**

An object of class data.frame with 65631 rows and 5 columns.

---

OrthG_Zf_Ch	<i>Orthologs genes database for mus zebrafish and chicken</i>
-------------	---

---

**Description**

Orthologs genes database for mus zebrafish and chicken

**Usage**

OrthG\_Zf\_Ch

**Format**

An object of class data.frame with 38394 rows and 5 columns.

---

 Plot\_MarkersHeatmap *Plot Markers in each cell type*


---

**Description**

This function integrate R package pheatmap to plot markers in each cell type

**Usage**

```
Plot_MarkersHeatmap(
  ConservedMarker,
  start_col = 2,
  module_colors = NA,
  heatmap_colors = NA,
  cluster_rows = FALSE,
  cluster_cols = FALSE,
  show_rownames = FALSE,
  show_colnames = FALSE,
  cellwidth = NA,
  cellheight = NA,
  legend = FALSE,
  annotation_legend = FALSE,
  annotation_names_row = FALSE,
  ...
)
```

**Arguments**

ConservedMarker	Markers table
start_col	numeric, indicating the start column of marker power in each cell type
module_colors	vector, indicating colors of modules (annotation_colors)
heatmap_colors	vector, indicating colors used in heatmap
cluster_rows	boolean values determining if rows should be clustered or hclust object
cluster_cols	boolean values determining if columns should be clustered or hclust object
show_rownames	boolean specifying if column names are be shown
show_colnames	boolean specifying if column names are be shown
cellwidth	individual cell width in points. If left as NA, then the values depend on the size of plotting window
cellheight	individual cell height in points. If left as NA, then the values depend on the size of plotting window
legend	logical to determine if legend should be drawn or not
annotation_legend	boolean value showing if the legend for annotation tracks should be drawn

annotation\_names\_row  
                    boolean value showing if the names for row annotation tracks should be drawn  
...                  parameter in pheatmap

**Value**

pheatmap object

**Examples**

```
data("pbmc_small")
all.markers <- Identify_Markers(pbmc_small)
all.markers <- Format_Markers_Frac(all.markers)
Plot_MarkersHeatmap(all.markers[,c(2,6,7,8)])
```

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