

# Package ‘snplinkage’

May 4, 2023

**Title** Single Nucleotide Polymorphisms Linkage Disequilibrium Visualizations

**Version** 1.0.0

**Description** Linkage disequilibrium visualizations of up to several hundreds of single nucleotide polymorphisms (SNPs), annotated with chromosomal positions and gene names. Two types of plots are available for small numbers of SNPs (<40) and for large numbers (tested up to 500). Both can be extended by combining other ggplots, e.g. association studies results, and functions enable to directly visualize the effect of SNP selection methods, as minor allele frequency filtering and TagSNP selection, with a second correlation heatmap. The SNPs correlations are computed on Genotype Data objects from the 'GWASTools' package using the 'SNPRelate' package, and the plots are customizable 'ggplot2' and 'gttable' objects and are annotated using the 'biomaRt' package. Usage is detailed in the vignette with example data and results from up to 500 SNPs of 1,200 scans are in Charlon T. (2019) <[doi:10.13097/archiveouverte/unige:161795](https://doi.org/10.13097/archiveouverte/unige:161795)>.

**Imports** gdsfmt, ggplot2, gtable, magrittr, stats, utils

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**Author** Thomas Charlon [aut, cre] (<<https://orcid.org/0000-0001-7497-0470>>),  
Karl Forner [aut],  
Alessandro Di Cara [aut],  
Jérôme Wojcik [aut]

**Maintainer** Thomas Charlon <charlon@protonmail.com>

**Repository** CRAN

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## R topics documented:

chisq_pvalues . . . . .	2
chisq_pvalues_gdata . . . . .	3
diamond_annot . . . . .	4
gdata_add_gene_annot . . . . .	5
gdata_add_gene_annot_aim_example . . . . .	5
gdata_add_gene_annot_hladr_example . . . . .	6
gdata_scans_annot . . . . .	6
gdata_snps_annot . . . . .	7
get_biomart_metadb . . . . .	7
ggplot_associations . . . . .	8
ggplot_ld . . . . .	9
ggplot_snp_pos . . . . .	9
gtable_ld . . . . .	10
gtable_ld_associations . . . . .	11
gtable_ld_associations_gdata . . . . .	12
gtable_ld_gdata . . . . .	13
load_gds_as_genotype_data . . . . .	14
parallel_apply . . . . .	14
print_qc_as_tex_table . . . . .	15
save_hgdp_as_gds . . . . .	16
select_region_idxs . . . . .	16
snprelate_allele_frequencies . . . . .	17
snprelate_ld . . . . .	17
snprelate_ld_select . . . . .	18
snprelate_qc . . . . .	19
%<>% . . . . .	20
%%% . . . . .	21
%>% . . . . .	21

## Index

22

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chisq_pvalues	<i>Compute Chi-squared p-values</i>
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---

### Description

Compute Chi-squared p-values

**Usage**

```
chisq_pvalues(
  m_data,
  response,
  adjust_method = "fdr",
  mlog10_transform = TRUE,
  n_cores = 1,
  ...
)
```

**Arguments**

m_data	Data matrix of observations by variables
response	Response vector of length the number of observations
adjust_method	Multiple testing p-value adjustment method. Passed to stats::p.adjust. 'fdr' by default.
mlog10_transform	Logical, transform p-values by minus log10. True by default.
n_cores	Number of cores
...	Passed to stats::chisq.test

**Value**

Chi-squared p-values

chisq\_pvalues\_gdata    *Compute Chi-squared p-values on a Genotype data object*

**Description**

Compute Chi-squared p-values on a Genotype data object

**Usage**

```
chisq_pvalues_gdata(
  gdata,
  snp_idxs,
  response_column = "region",
  response_value = "Europe",
  threshold = 2,
  ...
)
```

**Arguments**

<code>gdata</code>	Genotype data object
<code>snp_idxs</code>	SNPs indexes
<code>response_column</code>	Response column in gdata scans annotations data frame
<code>response_value</code>	Response value. The response vector will be a logical, true if equal to the value, false otherwise.
<code>threshold</code>	Keep only associations greater than the threshold
<code>...</code>	Passed to chisq_pvalues

**Value**

SNPs annotation data frame, chi-squared p-values in column pvalues

`diamond_annot`      *Get diamond ggplot layer.*

**Description**

Diamond ggplot layer for ggplot\_ld

**Usage**

```
diamond_annot(data, x = "x", y = "y", color = "color", size = 0.5)
```

**Arguments**

<code>data</code>	Data frame of 3 columns defining the diamonds
<code>x</code>	Name of the column for horizontal positions
<code>y</code>	Name of the column for vertical positions
<code>color</code>	Name of the column for color values
<code>size</code>	Radius of the diamonds

**Value**

gglayers

---

```
gdata_add_gene_annots  gdata_add_gene_annots
```

---

## Description

Add biomaRt gene annotations to Genotype Data object.

## Usage

```
gdata_add_gene_annots(  
  gdata,  
 .snp_idxs,  
  rsids_colname = "probe_id",  
  biomart_metadb = get_biomart_metadb()  
)
```

## Arguments

gdata	Genotype Data object
snp_idxs	SNP indexes
rsids_colname	Column of SNP annotation data frame with rs identifiers
biomart_metadb	List with slots snpmart and ensembl, corresponding to the biomart databases to query for SNP identifiers and gene names, respectively. See get_biomart_metadb function.

## Value

Genotype Data object

---

```
gdata_add_gene_annots_aim_example  
gdata_add_gene_annots_aim_example
```

---

## Description

Add ancestry informative markers gene annotations to Genotype Data object. Convenience function for the vignette to avoid querying biomaRt on build.

## Usage

```
gdata_add_gene_annots_aim_example(gdata, aim_idxs)
```

## Arguments

gdata	Genotype Data object
aim_idxs	AIM indexes in the example Genotype data object

**Value**

Genotype Data object

---

`gdata_add_gene_annots_hladr_example`  
*gdata\_add\_gene\_annots\_hladr\_example*

---

**Description**

Add HLA-DR gene annotations to Genotype Data object. Convenience function for the vignette to avoid querying biomaRt on build.

**Usage**

`gdata_add_gene_annots_hladr_example(gdata, hla_dr_idxs)`

**Arguments**

`gdata`            Genotype Data object  
`hla_dr_idxs`    HLA-DR indexes in the example Genotype data object

**Value**

Genotype Data object

---

`gdata_scans_annots`    *gdata\_scan\_annots*

---

**Description**

Get scans annotations from a Genotype Data object or a subset.

**Usage**

`gdata_scans_annots(gdata, scan_ids)`

**Arguments**

`gdata`            Genotype Data object  
`scan_ids`       Scan identifiers to subset

**Value**

Scans annotations data frame

---

gdata\_snps\_annot      *gdata\_snp\_annot*

---

### Description

Get SNPs annotations from a Genotype Data object or a subset.

### Usage

```
gdata_snps_annot(gdata,.snp_ids = NULL)
```

### Arguments

gdata	Genotype Data object
snp_ids	SNP identifiers to subset

### Value

SNP annotation data frame

---

get\_biomart\_metadb      *get\_biomart\_metadb*

---

### Description

To query gene names of SNPs, it is necessary to retrieve two objects using biomaRt::useMart. First, the object required to map SNP rs identifiers to ENSEMBL identifiers. Second, the object required to map ENSEMBL identifiers to common gene names. The function returns a list of two slots named snpmart and ensembl corresponding to each one, respectively. Once obtained it is saved to a local file.

### Usage

```
get_biomart_metadb(  
  filepath = extdata_filepath("bmart_meta.rds"),  
  host = "https://grch37.ensembl.org"  
)
```

### Arguments

filepath	Path to save the biomaRt objects
host	BiomaRt Ensembl host, by default https://grch37.ensembl.org

### Value

List of slots snpmart and ensembl as detailed above

`ggplot_associations`    *Ggplot associations*

---

## Description

Get SNPs associations ggplot, either as points or as a linked area. Optionally add labels to most associated points using ggrepel.

## Usage

```
ggplot_associations(
  df_snp,
  pvalue_colname = "pvalues",
  labels_colname = "probe_id",
  n_labels = 10,
  nudge = c(0, 1),
  linked_area = FALSE,
  byindex = linked_area,
  colors = if (linked_area) snp_position_colors(nrow(df_snp)) else "black"
)
```

## Arguments

<code>df_snp</code>	SNP annotation data frame with columns chromosome, position, and as specified by parameters <code>pvalue_colname</code> and optionally <code>labels_colname</code> .
<code>pvalue_colname</code>	Column name of <code>df_snp</code> with association values
<code>labels_colname</code>	Optional column name of <code>df_snp</code> with labels. Set to NULL to remove.
<code>n_labels</code>	Number of labels of most associated points to display.
<code>nudge</code>	Nudge parameter passed to <code>ggrepel::geom_label_repel</code> .
<code>linked_area</code>	Add a linked area to associations points, default FALSE
<code>byindex</code>	Display by SNP index or chromosomal position (default)
<code>colors</code>	Colors of SNPs

## Value

`ggplot`

---

<i>ggplot_ld</i>	<i>Ggplot linkage disequilibrium</i>
------------------	--------------------------------------

---

### Description

Display SNP r2 correlations using points or diamonds with text.

### Usage

```
ggplot_ld(
  df_ld,
  diamonds = length(unique(df_ld$SNP_A)) < 40,
  point_size = 120/sqrt(nrow(df_ld)),
  reverse = FALSE,
  reindex = TRUE
)
```

### Arguments

df_ld	Data frame with columns SNP_A, SNP_B, and R2. As returned by the snprelate_ld function.
diamonds	Should the values be displayed as diamonds or points ? Default is TRUE for less than 40 SNPs.
point_size	Size for geom_point. Ignored if diamonds is TRUE.
reverse	Reverse the display (horizontal symmetry)
reindex	If FALSE, SNPs are positionned following their IDs

### Value

ggplot

---

<i>ggplot_snp_pos</i>	<i>Ggplot SNPs position</i>
-----------------------	-----------------------------

---

### Description

Get SNPs position ggplot with mappings to combine with other ggplots. Optionally add labels and an upper subset.

### Usage

```
ggplot_snp_pos(
  df_snp,
  upper_subset = NULL,
  labels_colname = NULL,
  colors = snp_position_colors(nrow(df_snp))
)
```

**Arguments**

<code>df_snp</code>	SNP annotation data frame with a column named position and, if specified, one named as the <code>labels_colname</code> parameter.
<code>upper_subset</code>	Subset of <code>df_snp</code> for the positions on the upper side
<code>labels_colname</code>	Optional column name of <code>df_snp</code> to use as SNP labels.
<code>colors</code>	Colors for each SNP

**Value**

`ggplot`

`gtable_ld`

*Gtable of linkage disequilibrium and chromosomal positions*

**Description**

Creates a gtable of linkage disequilibrium and chromosomal positions ggplots. A `biplot_subset` parameter is available to add a second linkage disequilibrium ggplot to visualize the effect of a SNP selection.

**Usage**

```
gtable_ld(
  df_ld,
  df_snp,
  biplot_subset = NULL,
  labels_colname = NULL,
  diamonds = length(unique(df_ld$SNP_A)) < 40,
  point_size = ifelse(is.null(biplot_subset), 120, 80)/sqrt(nrow(df_ld)),
  title = "",
  title_biplot = "",
  ...
)
```

**Arguments**

<code>df_ld</code>	Data frame returned by <code>snpredict_ld</code>
<code>df_snp</code>	SNP annotations with columns <code>snpID</code> and <code>position</code>
<code>biplot_subset</code>	SNP indexes of the subset for the second ld plot
<code>labels_colname</code>	Column name of <code>df_snp</code> to use as SNP labels
<code>diamonds</code>	Display the values as diamonds or as points Default is TRUE for less than 40 SNPs.
<code>point_size</code>	Size for <code>geom_point</code> . Ignored if <code>diamonds</code> is TRUE.
<code>title</code>	Plot title
<code>title_biplot</code>	Optional biplot title
<code>...</code>	Passed to <code>ggplot_ld</code>

**Value**


---

gtable of ggplots

---

**gtable\_ld\_associations**

*Gtable of linkage disequilibrium and associations*

---

**Description**

Creates a gtable of a linkage disequilibrium, chromosomal positions, and association scores ggplots.

**Usage**

```
gtable_ld_associations(
  df_assocs,
  df_ld,
  pvalue_colname = "pvalues",
  labels_colname = "probe_id",
  n_labels = 5,
  diamonds = nrow(df_assocs) <= 40,
  linked_area = diamonds,
  point_size = 150/nrow(df_assocs),
  colors = snp_position_colors(nrow(df_assocs)),
  ...
)
```

**Arguments**

df_assocs	SNP annotation data frame with columns chromosome, position, and as specified by parameters pvalue_colname and optionally labels_colname.
df_ld	Data frame with columns SNP_A, SNP_B, and R2, as returned by the snprelate_ld function.
pvalue_colname	Column name of df.snp with association values
labels_colname	Optional column name of df.snp with labels. Set NULL to remove labels.
n_labels	Number of labels of most associated SNPs to display.
diamonds	Should the values be displayed as diamonds or points ? Default is TRUE for up to 40 SNPs.
linked_area	Add a linked area to associations points. Default same as diamonds.
point_size	Point size for ggplot_ld, ignored if diamonds is TRUE.
colors	Colors of SNPs
...	Passed to ggplot_associations

**Value**

gtable

---

**gtable\_ld\_associations\_gdata**

*Gtable of linkage disequilibrium and associations using a Genotype-Data object*

---

**Description**

Compute linkage disequilibrium using snprelate\_ld on the set of SNPs in the associations data frame and call gtable\_ld\_associations. Creates a gtable of a linkage disequilibrium, chromosomal positions, and association scores ggplots.

**Usage**

```
gtable_ld_associations_gdata(
  df_assocs,
  gdata,
  pvalue_colname = "pvalues",
  labels_colname = "probe_id",
  diamonds = nrow(df_assocs) <= 40,
  window = 15,
  ...
)
```

**Arguments**

<code>df_assocs</code>	SNP annotation data frame with columns chromosome, position, and as specified by parameters <code>pvalue_colname</code> and optionally <code>labels_colname</code> .
<code>gdata</code>	GenotypeData object, as returned by <code>load_gds_as_genotype_data</code>
<code>pvalue_colname</code>	Column name of <code>df.snp</code> with association values
<code>labels_colname</code>	Optional column name of <code>df.snp</code> with labels. Set NULL to remove labels.
<code>diamonds</code>	Should the values be displayed as diamonds or points ? Default is TRUE for up to 40 SNPs.
<code>window</code>	Window size for <code>snprelate_ld</code> . Forced to the total number of SNPs if <code>diamonds</code> is FALSE
<code>...</code>	Passed to <code>gtable_ld_associations</code>

**Value**

`gtable`

---

gtable_ld_gdata	<i>Gtable of linkage disequilibrium and positions using a GenotypeData object</i>
-----------------	---

---

## Description

Compute linkage disequilibrium using snprelate\_ld on a set of SNP indexes and call gtable\_ld. Two parameters are available to compute and compare minor allele frequency filtering and TagSNP selection by displaying two LD plots with their positions in the center. The maf and r2 parameters are used similarly and as follows: - compare baseline with MAF 5 gtable\_ld(gdata, snps\_idx, maf = 0.05) - compare baseline with TagSNP r2 = 0.8 gtable\_ld(gdata, snps\_idx, r2 = 0.8) - compare 5 gtable\_ld(gdata, snps\_idx, maf = c(0.05, 0.05), r2 = 0.8) - compare MAF 5 gtable\_ld(gdata, snps\_idx, maf = c(0.05, 0.1), r2 = c(0.8, 0.6))

## Usage

```
gtable_ld_gdata(
  gdata,
  snps_idx,
  maf = NULL,
  r2 = NULL,
  diamonds = length(snps_idx) < 40,
  window = 15,
  autotitle = TRUE,
  autotitle_bp = TRUE,
  double_title = FALSE,
  ...
)
```

## Arguments

gdata	GenotypeData object returned by load_gds_as_genotype_data
snps_idx	SNPs indexes to select
maf	Minor allele frequency threshold(s), see description
r2	TagSNP r2 threshold(s), see description
diamonds	Display the values as diamonds or as points Default is TRUE for less than 40 SNPs.
window	Window size for snprelate_ld. Forced to the total number of SNPs if diamonds is FALSE
autotitle	Set title to feature selection method(s), number of SNPs and chromosome
autotitle_bp	Set biplot title to feature selection method(s), number of SNPs and chromosome
double_title	Logical, if false (default) keep only biplot title
...	Passed to gtable_ld

**Value**


---

gttable of ggplots

---

`load_gds_as_genotype_data`

*Load GDS as Genotype Data*

---

**Description**

Open a connection to a snpgds file (cf. SNPRelate package) as a Genotype Data object.

**Usage**

```
load_gds_as_genotype_data(
  gds_file,
  read_snp_annot = TRUE,
  read_scan_annot = TRUE
)
```

**Arguments**

<code>gds_file</code>	Path of snpgds file
<code>read_snp_annot</code>	Read the SNPs' annotations
<code>read_scan_annot</code>	Read the scans' annotations

**Value**

Genotype Data object

---

`parallel_apply`

*Separate a matrix in a list of matrices of length the number of cores and apply a function on the columns in parallel*

---

**Description**

Separate a matrix in a list of matrices of length the number of cores and apply a function on the columns in parallel

**Usage**

```
parallel_apply(m_data, apply_fun, n_cores = 1, ...)
```

**Arguments**

m_data	Data matrix
apply_fun	Function to apply
n_cores	Number of cores
...	Passed to apply_fun

**Value**

apply\_fun return

---

`print_qc_as_tex_table` *print\_qc\_as\_tex\_table*

---

**Description**

Print information about quality control performed by the snprelate\_qc function.

**Usage**

```
print_qc_as_tex_table(  
  gdata_qc,  
  label = "qc",  
  caption = paste("Quality control and feature selection of the subset of the",  
    "human genome diversity project dataset."  
)
```

**Arguments**

gdata_qc	Genotype Data object object returned by snprelate_qc
label	Label of the Tex table
caption	Caption of the Tex table

**Value**

Prints knitr::kable object using cat

`save_hgdp_as_gds`      *save\_hgdp\_as\_gds*

### Description

Save the HGDP SNP data text file as a Genomic Data Structure file

### Usage

```
save_hgdp_as_gds(paths = hgdp_filepaths(), outpath = tempfile(), ...)
```

### Arguments

<code>paths</code>	Paths of the zip, txt, and gds files
<code>outpath</code>	Output GDS file path
<code>...</code>	Passed to save_genotype_data_as_gds

### Value

Path of the saved gds file

`select_region_idxs`      *select\_region\_idxs*

### Description

Select SNP indexes corresponding to a specific genomic region.

### Usage

```
select_region_idxs(
  gdata,
  chromosome,
  position_min = -Inf,
  position_max = Inf,
  n_snps = 0,
  offset = 0
)
```

### Arguments

<code>gdata</code>	Genotype Data object
<code>chromosome</code>	Chromosome to select
<code>position_min</code>	Minimum base pair position to select
<code>position_max</code>	Maximum base pair position to select
<code>n_snps</code>	Maximum number of SNPs to return
<code>offset</code>	Number of SNPs to offset

**Value**

SNP indexes of Genotype Data object

---

`snprelate_allele_frequencies`

*Compute allele frequency and snp missing rate*

---

**Description**

Wrapper over SNPRelate::snpgdsSNPRateFreq

**Usage**

```
snprelate_allele_frequencies(
  gdata,
  snps_idx = NULL,
  scans_idx = NULL,
  quiet = FALSE
)
```

**Arguments**

<code>gdata</code>	A GenotypeData object
<code>snps_idx</code>	Vector of snps indices
<code>scans_idx</code>	Vector of scans indices
<code>quiet</code>	Whether to be quiet

**Value**

A data frame of `snps_idx`, `snps_ids`, `allele1`, `allele2`, `maf`, `missing` where `allele1` and `allele2` are the rates of the alleles, and `maf` the minimum of the 2. `Missing` is the missing rate. N.B: the allele rates are computed on the non missing genotypes, i.e. their sum equals 1.

---

`snprelate_ld`

*Wrapper for snpgdsLDMat to compute r2*

---

**Description**

Wrapper for snpgdsLDMat to compute r2

**Usage**

```
snprelate_ld(
  gdata,
  window_size = 0,
  min_r2 = 0,
  snps_idx = NULL,
  scans_idx = NULL,
  threads = 1,
  quiet = FALSE
)
```

**Arguments**

gdata	A GenotypeData object
window_size	Max number of SNPs in LD window, 0 for no window
min_r2	Minimum r2 value to report
snps_idx	Indices of snps to use
scans_idx	Indices of scans to use
threads	The number of threads to use
quiet	Whether to be quiet

**Value**

A data frame with columns SNP\_A, SNP\_B, R2 for  $r2 \geq \text{min\_r2}$

snprelate\_ld\_select    *Wrapper for snpgdsLDpruning to select Tag SNPs*

**Description**

The tagged SNP set is (by sliding window) representative and strongly not redundant.

**Usage**

```
snprelate_ld_select(
  gdata,
  window_length = 500L,
  min_r2,
  window_size = NA,
  snps_idx = NULL,
  scans_idx = NULL,
  remove.monosnp = FALSE,
  autosome.only = FALSE,
  method = "r",
  threads = 1,
```

```
quiet = FALSE,
...
)
```

**Arguments**

gdata	A GenotypeData object
window_length	Max length in kb of the window
min_r2	Minimum r2 value to report
window_size	Max number of SNPs in LD window
snps_idx	Indices of snps to use
scans_idx	Indices of scans to use
remove.monosnp	if TRUE, remove monomorphic SNPs
autosome.only	if TRUE, use autosomal SNPs only; if it is a numeric or character value, keep SNPs according to the specified chromosome
method	"composite", "r", "dprime", "corr", see details
threads	The number of threads to use, currently ignored
quiet	Whether to be quiet
...	Forwarded to SNPRelate::snpgdsLDpruning

**Value**

A list of SNP IDs stratified by chromosomes.

`snprelate_qc`

`snprelate_qc`

**Description**

Quality control using SNPRelate functions.

**Usage**

```
snprelate_qc(
  gdata,
  samples_nas = 0.03,
  ibs = 0.99,
  keep_ids = NULL,
  snps_nas = 0.01,
  maf = 0.05,
  tagsnp = 0.8,
  n_cores = 1
)
```

**Arguments**

gdata	Genotype data object
samples_nas	NA threshold for samples, default 3 pct
ibs	Samples identity by state threshold, default 99 pct
keep_ids	Samples ids to keep even if IBS is higher than threshold. Used for monozygotic twins.
snps_nas	NA threshold for SNPs, default 1 pct
maf	Minor allele frequency threshold, default 5 pct
tagsnp	TagSNP r2 correlation threshold, default 0.8
n_cores	Number of cores

**Value**

List of gdata, Genotype data object, and df\_qc, QC info data frame

%&lt;&gt;%

*Assignment pipe***Description**

Pipe an object forward into a function or call expression and update the ‘lhs’ object with the resulting value. Magrittr imported function, see details and examples in the magrittr package.

**Arguments**

lhs	An object which serves both as the initial value and as target.
rhs	a function call using the magrittr semantics.

**Value**

None, used to update the value of lhs.

---

%\$%

*Exposition pipe*

---

### Description

Expose the names in ‘lhs’ to the ‘rhs’ expression. Magrittr imported function, see details and examples in the magrittr package.

### Arguments

lhs	A list, environment, or a data.frame.
rhs	An expression where the names in lhs is available.

### Value

Result of rhs applied to one or several names of lhs.

---

%>%

*Pipe*

---

### Description

Pipe an object forward into a function or call expression. Magrittr imported function, see details and examples in the magrittr package.

### Arguments

lhs	A value or the magrittr placeholder.
rhs	A function call using the magrittr semantics.

### Value

Result of rhs applied to lhs, see details in magrittr package.

# Index

%<>%, 20  
%>%, 21  
%\$%, 21

chisq\_pvalues, 2  
chisq\_pvalues\_gdata, 3

diamond\_annots, 4

gdata\_add\_gene\_annots, 5  
gdata\_add\_gene\_annots\_aim\_example, 5  
gdata\_add\_gene\_annots\_hladr\_example, 6  
gdata\_scans\_annots, 6  
gdata\_snps\_annots, 7  
get\_biomart\_metadb, 7  
ggplot\_associations, 8  
ggplot\_ld, 9  
ggplot\_snp\_pos, 9  
gttable\_ld, 10  
gttable\_ld\_associations, 11  
gttable\_ld\_associations\_gdata, 12  
gttable\_ld\_gdata, 13

load\_gds\_as\_genotype\_data, 14

parallel\_apply, 14  
print\_qc\_as\_tex\_table, 15

save\_hgdp\_as\_gds, 16  
select\_region\_idxs, 16  
snprelate\_allele\_frequencies, 17  
snprelate\_ld, 17  
snprelate\_ld\_select, 18  
snprelate\_qc, 19